





1.6. Types of columns

		
Column type	wide bore	spin
Order No.:	IC-C-05-25	IC-M-05-50
Package size:	25 units / pack	50 units / pack
Elution volume	3 mL = 1 mL + 2 mL	500 µL = 200 µL + 300 µL
Recommended loading:	< 300 ng	< 100 ng

Use of adapters and reservoirs for loading recommended (Order no.: LB-08-13)

1.7. Preparation

1.7.1. Cleaning

All equipment has to be clean and not contaminated with materials that might cause interference with the analysis. All equipment should be washed with a mild detergent solution and then rinsed thoroughly with purified water. This includes glass ware, adapters and syringe barrels used for sample reservoirs. In between assays it is sufficient to rinse with methanol and water. This helps to prevent cross-contamination of samples.

1.7.2. Preparation of reagents

Prepare solutions every week or as needed.

CAUTION: Methanol, Acetonitrile and the solutions made thereof are flammable. Keep containers in a safe place and tightly capped when not in use.

Extraction solvent:

Methanol/Water

Use Methanol HPLC grade only. Use 800 mL methanol and 200 mL deionized water.

Methanol/PBS

Use Methanol HPLC grade only. Use 700 mL methanol and 300 mL PBS buffer, mix.

Diluting Buffer: Bicarbonate solution

2.5 g NaCl, 0.5 g NaHCO₃ bring to 100 mL with deionized water.

Diluting Buffer: PBS

8.0 g NaCl, 1.2 g Na₂HPO₄, 0.2 g KH₂PO₄, 0.2 g KCl, dissolve in approximately 990 mL purified water, adjust pH to 7.0 with concentrated HCl, bring to 1 liter with purified water.

Wash Buffer: PBS/Methanol

Use 100 mL methanol and 900 mL PBS, mix.

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Subject to change without notice.

aokin AG 07/2020

FB-593-05-R4



Methanol / 1 % acetic acid for elution

Use HPLC Grade methanol only.

Use 99 mL methanol and 1 mL acetic acid, mix.

1.8. Materials required for the sample preparation and the HPLC

<i>aokinImmunoClean C FUM</i>	(IC-C-05-25, 25 units/pack)
<i>aokinImmunoClean M FUM</i>	(IC-M-05-50, 50 units/pack)
<i>aokin</i> Filter Paper	(LB-05-10-100, 100 units/pack)
Glass fiber filters GF/F	(LB-04-13-GF/F-100, 100 units/pack)
Reaction tubes (2 ml, with lid)	(LB-05-05, 500 units/pack)
Test tubes (15 ml, with lid)	(LB-05-01-100, 100 units/pack)
Test tubes (50 ml, with lid)	(LB-05-02-250, 250 units/pack)
Methanol, HPLC Grade	(LB-03-02-1000, 1 L)
Sodium Chloride, pure	
Acetonitrile, HPLC Grade	
Distilled, reverse osmosis or deionized water	
Graduated cylinder stand (50 mL)	(LB-08-16, 1 unit)
Graduated cylinder stand (250 mL)	(LB-08-17, 1 unit)
Cuvette Rack	(LB-05-04)
Digital Scale	(LB-07-04, 1 unit)
Commercial blender, with metal or glass beaker for use with Acetonitrile mixtures	(EX-08, 1 unit)
Commercial blender, with plastic beaker (200 mL) for use with Methanol mixtures	(EX-07-06, 1 unit)
Vacuum-pump (diaphragm pump)	(LB-04-10, 1 unit)
Trap for Vacuum-pump (vacuum bottle), 500 mL	(LB-04-12, 1 unit)
Vacuum manifold	(LB-04-09, 1 unit)
Filter funnel (for retaining paper filters)	(LB-06-01, 1 unit)
Adjustable Micropipette, 1000 µL	(LB-04-05-1000, 1 unit)
Micropipette tips for adjustable Micropipette, 1000 µL	(LB-04-08-1000L, 500 units/pack)
<i>aokinReferenceMatrixMaterial FUM</i>	(RMM-05)

1.9. Set up and equilibration of columns

Allow column to be at ambient temperature. Remove bottom cap and place the column onto a vacuum manifold, or in a pump stand or collection tube. Open top cap and fill column with PBS. Connect adapter and a reservoir to the column. Use a flow rate of 1 mL/min and have 1-2 ml pass through the column. This step ensures an equilibration of the column. Close the valve again to stop the flow.

2. Points of critical importance for reproducibility and recovery

2.1. Representative sampling

A representative sample is essential for accurate and reliable results. Samples should be collected and ground before taking a subsample. Contamination of mycotoxin may differ significantly within a single batch and from kernel to kernel.

2.2. Sample preparation

Different procedures require different reagents. Please make sure that your protocol consists of the following points:

- Dilute filtrate to 10 % solvent ratio
- Adjust to neutral pH.
- Remove all precipitation by glassfiber filtration using a 1.7 µm mesh size.
- Equilibrate column to room temperature, best by rinsing with PBS.
- Load column with flow rate of 1 mL/min.
- Wash column with PBS, deionized water is not recommended.
- Dry column by vacuum or air pressure.
- 1 mL Apply Methanol/1%Acetic Acid. Incubate for 3 minutes by stopping flow. Apply 2 mL Methanol/1%Acetic Acid.
- Elute by vacuum or air pressure at 1 mL/minute or by back flushing with a syringe.
- Quantify the concentration by comparing the sample peak height or area to the standard.

aokinImmunoClean FUM columns have been optimized for quantitative measurement of Fumonisin in many commodities. Test methods vary in the amount of sample passed through the affinity column resulting in different limits of detection.

General recommendation:

- Perform test from beginning to end without interruptions.
- Load sample on column immediately after filtration.
- Mix the eluate in the cuvette very well before injecting eluate into HPLC.
- Avoid contact of any test reagents or solutions (such as acetonitrile, methanol or column eluate) with rubber or soft flexible plastic. These materials may leach chemicals into the sample.
- Maintain a slow and steady flow rate through the column during sample loading.
- Elute the column slowly, do an incubation step.

Example Procedures:

A1. Grain and corn

Sample extraction:

- Weigh 50 g ground sample and place in blender jar. Add 5 g NaCl. Use 200 mL beaker for best blending.
- Add 100 mL Methanol/water (80/20) or Methanol/buffer (70/30)
- Cover beaker and blend at high speed for 3 minutes.
- Remove cover from beaker and pour extract into fluted filter paper. Collect filtrate in a clean vessel.
- Transfer 10 mL filtered extract into another clean vessel.
- Dilute extract with 40 mL PBS. Mix well.
- Check pH to be neutral, if required neutralize.
- Use a glass fiber filtration through 1.7 µm glass microfibre filter into a clean vessel.

B. Set up column

- Connect **aokinCA**adapter and a 20 mL syringe barrel (best flow when bubble free).
- Place on vacuum manifold or pump stand.
- Flush with 2 mL PBS

Column Chromatography:

- Pass 10 mL filtered diluted extract through **aokinImmunoClean** column at a rate of about 1mL/minute (about 1 drop/second) until air comes through column.
- Pass 10 mL of PBS through the column at a rate 3 mL/minute.
- Repeat if column bed is dark. Dry column with air flow.
- Place new collection tube under the **aokinImmunoClean** column.

Elution:

- Add 1 mL Methanol/1%Acetic Acid Incubate for 5 minutes by stopping flow.
- Pass additional 2 mL Methanol/1%Acetic Acid through **aokinImmunoClean** at a rate of 1 drop/second.
- Centrifuge eluate at 15.000 g to remove precipitation or alternatively add 1.5 mL water to dissolve precipitation.

C. Recovery

- Recovery of > 80% tested in PBS buffer.
- Exact results are found in the attached data sheet.
- Test the recovery of **aokinImmunoClean** columns with your protocol and HPLC technique, and use a correction factor as determined.

D. HPLC setup

Derivatization and fluorescence detection or alternative mass spectrometer detection recommended.



A2. Beer

Sample extraction:

- Use 5 mL beer

B. Set up column

- Connect **aokin**CA*adapter* and a 20 mL syringe barrel (best flow when bubble free).
- Place on vacuum manifold or pump stand.
- Flush with 2 mL PBS

Column Chromatography:

- Pass 5 mL beer through **aokin**ImmunoClean column at a rate of about 1mL/minute (about 1 drop/second) until air comes through column.
- Pass 10 mL of Bicarbonate solution through the column at a rate 3 mL/minute.
- Dry column with air flow.
- Place new collection tube under the **aokin**ImmunoClean column.

Elution:

- Add 1 mL Methanol/1%Acetic Acid. Incubate for 5 minutes by stopping flow.
- Pass additional 2 mL Methanol/1%Acetic Acid through **aokin**ImmunoClean at a rate of 1 drop/second.
- Centrifuge eluate at 15.000 g to remove precipitation or alternatively add 1.5 mL water to dissolve precipitation.

C. Recovery

- Recovery of > 80 % tested in PBS buffer.
- Exact results are found in the attached data sheet.
- Test the recovery of **aokin**ImmunoClean columns with your protocol and HPLC technique, and use a correction factor as determined.

D. HPLC setup

Derivatization and fluorescence detection or alternative mass spectrometer detection recommended.

Liabilities

The customer is solely and fully responsible for educating oneself about the proper testing and sampling procedures using this product. **aokin** makes no warranty of any kind. **aokin** is not liable or responsible for any unsatisfactory or faulty results.

Methods and references

AOAC method 2001.04 Determination of Fuminisin B1 and B2 in Corn and Cornflakes by Liquid Chromatography and Immunoaffinity Column Clean Up J. of AOAC Int. 84(6) 1828-1837, 2001

CEN 14352 Determination of Fumonisin B1 and B2 in maize based foods – HPLC method with immunoaffinity column clean up.

CEN 16187 Determination of Fumonisin B1 and B2 in processed maize containing foods for infants and young children – HPLC method with immunoaffinity column clean up and fluorescence detection after precolumn derivatization.

Ordering and technical support

To place an order please contact **aokin** at orders@aokin.com

For technical information please contact service@aokin.com

Please contact the application laboratory and service staff for all questions relating to the optimal use of our columns We will be glad to assist you.