

INSTRUCTIONS FOR USE

aokin mycontrol **T2/HT2**

Order No.: MY-QC-78

Sample preparation with aokinQuickClean columns (SPE)



Analytical-kit for rapid and quantitative determination of T-2- and HT-2-Toxin (T2/HT2).

Materials

aokinmycontrolT2/HT2 (Order No.: MY-QC-78-100)

Package content

A) Materials for sample preparation:

aokinExtractionSolventT2/HT2, Extraction solution
aokinExtractionSalt T2/HT2 + spoon
aokinQuickClean T2/HT2, centrifuge columns
aokinmycontrol T2/HT2 Precipitation buffer
(transparent cap)

Filter paper

Reaction tubes 2 mL

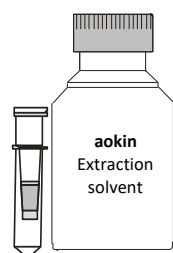


Figure 1: aokinQuickClean column with reaction tube and Extraction solvent (1 L bottle)

B) Materials for analytical measurement:

aokinReactionBuffer, Reaction buffer
aokinmycontrol T2/HT2, Reagent 1 (yellow cap),
F-T2/HT2, (for 5 analyses each)
aokinmycontrol T2/HT2, Reagent 2 (black cap),
A-T2/HT2, (for 5 analyses each)

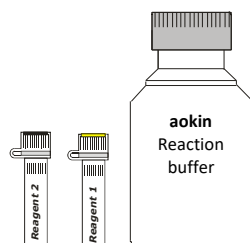


Figure 2: Reagent 1, F-T2/HT2 (yellow cap), Reagent 2, A-T2/HT2 (black cap) and Reaction buffer (1 L bottle)

C) Materials for internal quality control:

aokinmycontrol T2/HT2, negative control T2/HT2
(transparent), for zero value measurements
aokinmycontrol T2/HT2, Reagent 1 (yellow cap),
F-T2/HT2, (for 5 analyses each)
aokinmycontrol T2/HT2, Reagent 2 (black cap),
A-T2/HT2, (for 5 analyses each)

Note: All substances provided are precisely weighed and calibrated. Control of the volume and concentration of the individual solutions are essential for the precision of the analysis.

Caution: The extraction solvent may contain methanol. Work with professional care.

Storage Conditions: Reagents 1 and 2 must be stored at temperature of +2 - +10°C. All other components may be stored at room temperature.

Quality Control: All materials and reagents are prepared according to strict quality control protocols. Exchanging reagents between kits having different Lot-numbers will lead to erroneous results and is not permitted.

Order Information:

aokinmycontrol T2/HT2 (Order No.: MY-QC-78-100)

Introduction

aokinmycontrolT2/HT2 is a rapid and precise quantitative method for analyzing of T-2 and HT-2-Toxin (T2/HT2). It has been specifically designed and calibrated for the analysis of wheat and includes a sample preparation with solid phase extraction (SPE) columns. Samples in the µg/kg range (ppb = parts per billion range) can be analysed for T-2 and HT-2-Toxin in 15 minutes.

aokinmycontrolT2/HT2 is available with a calibration, which has been validated for wheat. Please use professional care and check the accuracy by regularly analyzing reference materials (e.g. aokinReferenceMatrix Materials) and/or standards. Participation in proficiency tests is recommended.

aokin will gladly assist you customising the test for your specific sample type and application. Please do not hesitate to contact us.

Sample	wheat
Time required for sample preparation	12 minutes
Time required for measurement	3 minutes
Analysis	
	Measurement range [µg/kg]
Range 1	70 – 420
Range 2	140 – 840
Range 3	280 – 1680

T-2 and HT-2 Toxins

T-2 and HT-2 are mycotoxins. They naturally occur in molds by *Fusarium sp.* fungus. It is toxic to humans and animals. As a consequence, it is strongly recommended to monitor the content in grain and corn food and feed raw materials and products.

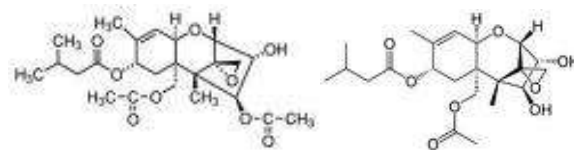


Figure 3: Chemical formula for T-2 Toxin (C₂₄H₃₄O₉ molecular weight: 466,52 g/mol) and HT-2 Toxin (C₂₂H₃₂O₈; molecular weight: 424,48 g/mol)

Recommended Accessories

All required materials are available from aokin.

Tel.: +49 30 9489 2160

	Order No.:
aokinextractor (food blender)	EX-07-06
aokinwatchbox (timer for food blender)	EX-07-06-4
Weighing scale, d = 0,01 g	LB-03-04
Eppendorf centrifuge, variable g-force	LB-04-04
Variable pipettes (1000 µl)	LB-04-05-1000
Pipette tips (1000 µl)	LB-04-08-1000
Funnels	LB-05-04
Dispensette	LB-08-01

the filter paper and filter cake. Shake/stir the filtrate to ensure homogeneity.

Sample preparation

The following protocol is an example. The quantification ranges are dependent on dilutions. Actual volume settings in the software may vary.

Note: It is of critical importance to use the correct sample preparation protocol for each determination. Use volumes displayed in the *aokin* software.

1. Quality control

Included in the analytical kit there are following additional materials for your internal quality control: **Reagent 1**, **Reagent 2**, negative control samples (labelled **negative control**, corresponding to samples free of mycotoxin) and a positive control sample. Please perform measurements of negative controls regularly, this ensures the accuracy of your determinations. If you notice increased values, change cuvette and repeat measurement. If sample results remain high, contact the *aokin* team.

2. Sample collection, grinding and mixing

The analysis sample is collected, ground, and homogenised according to an approved procedure. Small samples may be ground using the *aokinextractor*.

3. Weighing and extraction

Weigh 15 g of your sample, add one spoon (1,5 g) of *aokinExtractionSalt* T2/HT2 and 30,4 g extraction solution (35 ml *aokinExtractionSolvent* T2/HT2 at 20°C) directly into the extraction beaker (Figure 4). Preferentially the exact volume is applied using a dispenseette.



Figure 4: Weighing

Close the extraction beaker with the lid (with the blending knives). Blend for 3.5 minutes. The recommended protocol has blending times alternating with resting time to avoid heating of the sample and is as follows: mix for 30 seconds, pause for 1 minute, mix for 30 seconds and so on (until 3.5 minutes of blending time).

Use the *aokinwatchbox* (a preprogrammed timer) to conveniently and automatically complete this extraction protocol.



Figure 5: Extracting with the *aokinextractor* (blender)

3. Filtration

Place the filter on a suitable funnel and the funnel onto a collection container. Open the extraction beaker and pour the contents over the filter and collect the filtrate. Discard



Figure 6: Filtration

4. Use of *aokinQuickClean* column

Place an *aokinQuickClean* **T2/HT2** column in a collection tube and add 900 µl of the filtrate (Figure 7). Place it in the centrifuge and spin for 3 minutes at 5.000 x g. Place second *aokinQuickClean* **T2/HT2** column in a collection tube and add ≥ 650 µL of the purified filtrate. Place it in the centrifuge and spin for 3 minutes at 5000 x g.

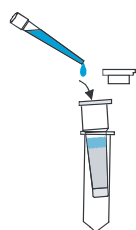


Figure 7: Pipetting of the extract onto the *aokinQuickClean*T2/HT2 column

5. Precipitation

Add 400 µl of column-filtrate into the *aokinmycontrol* T2/HT2 precipitation buffer (transparent cap) and mix it well. In case a precipitation is visible centrifuge with maximum g-force (> 10.000 x g) for 5 minutes.

Transfer the supernatant into a clean tube. Your sample is now ready for analysis.

6. Analyzing

Use supernatant of precipitation for analyzing in the *aokinspectrometerFP470*. Please follow detailed instructions for spectrometer use (*aokinspectrometerFP470* & *aokinLHW03* Instructions for use).

This includes:

- 1) Place *Reagents 1* and *2* into position A6 and B6 of the sample rack of your spectrometer.
- 2) Place a 25 ml *Clean1*-glass container, filled with *Clean1* solution into the *Clean1*-position, on the left side, next to the palette.
- 3) Place a 25 ml *neg. control* T2/HT2-glass container, filled with negative control T2/HT2 solution = 4 mL *aokinExtraction Solvent* T2/HT2 + 8 mL *aokinmycontrol* T2/HT2 precipitation buffer in the *Clean2*-position, on the left side of the palette.
- 4) Place an empty 2 mL vial in position A1 of the palette.
- 5) Place an empty waste bottle in the holder. Check presence of Reaction buffer and check if tubing is below the surface.
- 6) Place a new cuvette with a clean stirrer into the spectrometer.

7. Quality control

Included in the analytical kit there are following additional materials for your internal quality control: Reagent 1, Reagent 2, negative control samples (labelled negative control, corresponding to samples free of mycotoxin) and a positive control sample.

If problem persists calibrate. Please contact the aokin team for any support needed.

Please perform measurements of negative controls regularly, this ensures the accuracy of your determinations.

If you notice increased values, change cuvette and repeat measurement. If results remain high perform an offset correction of the calibration based on the negative control results. In addition, the use of recovery corrections preferentially by using sample extracts from certified reference matrix samples or alternatively based on the positive controls included in the kit.

Conversion factor: analyte concentration in cuvette (nM) to amount in sample (µg/kg)

aokinmycontrol T2/HT2
Standard

Step 1: Extraction

- Sample mass: $m_{\text{Sample}} = 15 \text{ g}$
- Volume extraction solvent: $V_{\text{Extraction solvent}} = 35 \text{ ml}$
- Molar mass T2: $MW_{T2} = 445,555 \left[\frac{\text{g}}{\text{mol}} \right]$

Mycotoxin concentration in the sample extract:

$$c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}} = \frac{m_{\text{Sample}} [\text{kg}]}{V_{\text{Solvent}} [\text{l}] * MW_{\text{Mykotoxin}} \left[\frac{\text{g}}{\text{mol}} \right]} * c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}} = \frac{0.015}{0.035 * 445,555} * c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}} = 0.00096188 * c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}}$$

Step 2: Purification
with **aokin QC T2/HT2**

- Volume sample extract load to the **aokin QC** column: $V_{\text{loaded sample extract}} = 0,9 \text{ ml}$
- Volume eluate from the **aokin QC** column: $V_{\text{elute}} = 0,9 \text{ ml}$

Mycotoxin concentration in the column eluate:

$$c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Eluate}} = \frac{V_{\text{load}} [\text{ml}]}{V_{\text{elute}} [\text{ml}]} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}} = \frac{0,9}{0,9} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}} = 1 * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}}$$

Step 3: Dilution

- Volume Eluate: $V_{\text{eluate}} = 0,4 \text{ ml}$
- Total volume: $V_{\text{total}} = 1,2 \text{ ml}$

$$c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Diluted}} = \frac{V_{\text{eluate}} [\text{ml}]}{V_{\text{total}} [\text{ml}]} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}} = \frac{0,4}{1,2} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}} = 0,333 * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Extract}}$$

Step 4: Measurement
aokin FP 470 / LHW 03

- Sample volume: $V_{\text{column eluate}} = V_{\text{sample}} = 600 \mu\text{l}$
- Total volume in the cuvette: $V_{\text{cuvette}} = 2600 \mu\text{l}$

Mycotoxin concentration in the cuvette:

$$c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Cuvette}} = \frac{V_{\text{Sample}} [\mu\text{l}]}{V_{\text{cuvette}} [\mu\text{l}]} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Eluate}} = \frac{600}{2600} * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Eluate}} = 0.23 * c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Eluate}}$$

Conversion factor: Extraction, Purification and Measurement
It follows from 1 to 4 above the conversion factor

$$c \left[\frac{\mu\text{mol}}{\text{l}} \right]_{\text{Cuvette}} = 0,00096188 * 1 * 0,333 * 0.23 * c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}} = 0.00007367 * c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}} \quad \text{or}$$

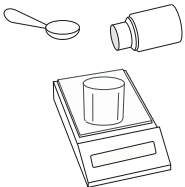
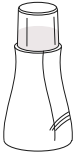

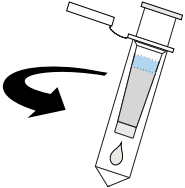
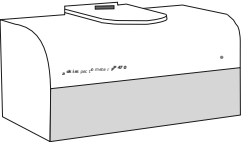
$$c \left[\frac{\mu\text{g}}{\text{kg}} \right]_{\text{Sample}} = \frac{1}{0.07367} * c \left[\frac{\text{nmol}}{\text{l}} \right]_{\text{Cuvette}} = 13.515 * c \left[\frac{\text{nmol}}{\text{l}} \right]_{\text{Cuvette}}$$

T-2/HT-2 toxin / for wheat flour samples

 **aokinmycontrol T2/HT2**

Procedure:

Standard

Extraction		Weighing: 15 g sample 1,5 g aokinExtractionSalt T2/HT2 35 mL aokinExtractionSolvent T2/HT2
		Extraction: 3,5 min mixing with aokinwatchbox
		Filtration: collect filtrate (discard filter cake)
Purification		SPE-Filtration: 1. column: 900 µL filtrate on aokinQuickClean column 3 min centrifuge at 5000 x g 2. column: ≥ 650 µL filtrate of the 1 st aokinQuickClean on 2 nd aokinQuickClean column 3 min centrifuge at 5000 x g, use second column filtrate for measurement
		Precipitation: 400 µL column filtrate into <i>Precipitation buffer</i> (transparent cap) 5 min centrifuge at > 10.000 x g transfer supernatant into clean 2 mL reaction tube
Measurement		Automatic Analyse (FP470 / LHW03): place the 2ml reaction tube in the sample holder of the <i>LHW03</i> 1800 µL aokin Reaction buffer 600 µL sample (diluted 1:1 - RANGE 1) (diluted 1:2 - RANGE 2) (diluted 1:4 - RANGE 3) 100 µL aokinmycontrol T2/HT2 Reagent 1

100 µL **aokinmycontrol T2/HT2** Reagent 2

T2/HT2 = T2/HT2 Toxins, Conversion factor: 1 nmol T2/HT2/l in cuvette = 13,515 µg/kg in sample